

Report:

Water quality data from monthly sampling at Bass Lake



Photo credit: Bass Lake Property Owners Association

Prepared for:
Bass Lake Conservancy District
Knox, IN

Prepared by:
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Report Date: July 9, 2026



1. Summary

Aquatic Control completed data and sample collection at Bass Lake on June 25, 2026. Three open-water sites were sampled for water quality (see map and coordinates on page 2). At each of these three sites, a YSI ProDSS Multi-parameter water quality meter and ProDSS 4-port sensor cable were used to measure vertical profiles of pH, dissolved oxygen (DO), temperature, conductivity, phycocyanin, and chlorophyll-*a* at 1-meter intervals through the water column. Secchi depth was also measured at each sampling site. Water samples were collected at the surface and bottom of each site and overnight shipped to Aquatic Control's laboratory. Surface water samples were analyzed for total phosphorus, total nitrogen, total Kjeldahl nitrogen, and chlorophyll-*a* (via extraction and fluorescence). Bottom water samples were analyzed for total phosphorus and soluble reactive phosphorus. A sample was also collected from the groundwater pump house outflow for analysis of pH, DO, temperature, conductivity, phycocyanin, and chlorophyll-*a* using the multi-parameter sonde, while water samples were analyzed in the laboratory for total phosphorus, total nitrogen, total Kjeldahl nitrogen, and chlorophyll-*a* (via extraction and fluorescence). Five near-shoreline sites were also visited within the lake to collect water samples for analysis of algal identification and enumeration in the laboratory.

At the time of sampling June 25, 2026, Bass Lake was stratified based on dissolved oxygen and temperature data. pH, conductivity, and dissolved oxygen were all within normal ranges for a freshwater lake in the summer. Algal pigments indicative of planktonic algae or cyanobacteria were also relatively low in the open waters. Phosphorus and nitrogen concentrations were moderate to high, in line with a eutrophic (biologically productive) lake. Secchi depths at all three sampling sites had improved slightly relative to measurements from May. Samples collected from shoreline sites for algae identification and enumeration showed high densities of cyanobacteria, indicating that there is likely more phosphorus available in those areas for cycling by plants and algae. Toxin testing was not performed on these samples but can be added in the future if desired by Bass Lake. Sampling will continue monthly through September 2026.

2. Sampling Site Information



Sample Site Coordinates

Sample Site	Latitude	Longitude
Site 1- Water Quality	41.2136	-86.5942
Site 2- Water Quality	41.2251	-86.5836
Site 3- Water Quality	41.2357	-86.5765
Pumphouse Outflow	41.2298	-86.5759
Site 1- Algae ID&E	41.2120	-86.6113
Site 2- Algae ID&E	41.2242	-86.6015
Site 3- Algae ID&E	41.2216	-86.5791
Site 4- Algae ID&E	41.2312	-86.5867
Site 5- Algae ID&E	41.2409	-86.5742

3. Results

3.1 pH

pH is the measure of how acidic or basic water is, reported in standard units (S.U.). A pH of 7 is neutral, and a range of 6-9 is typical for freshwater. A pH range of 6.5-9 is required for health and protection of aquatic organisms (i.e., survival, growth, and reproduction).

Results:

pH levels measured in the open water sites at Bass Lake ranged from about 8.80 (highest value measured) at the water surface down to about 6.35 (lowest value measured at the bottom of the water column). pH in the pumphouse water was 7.40 (see Table 1 and Figure 1 for results). Therefore, pH levels in Bass Lake were on the higher end of normal range at the water surface, and the groundwater flowing into the lake was closer to neutral. Common factors that contribute to higher pH in lakes can include high photosynthetic activity by plants and algae (including algae blooms), and relatively higher alkalinity or carbonate geology in the lake or watershed. Higher pH levels are not inherently a problem or unusual but can be an indicator of eutrophication if levels are higher in the afternoon and lower in the early morning.

3.2 Dissolved Oxygen

Dissolved oxygen (DO) is the amount of oxygen gas that is mixed into water and available for aquatic organisms to breathe. DO is considered an important metric for water quality since it is essential for survival of aquatic life. Changes in DO can occur for several reasons including changes in water temperature, wind action, and biomass of plants and algae in a lake. When plants and algae respire overnight, this process consumes oxygen. When these organisms are conducting photosynthesis during the daytime, this process produces oxygen. Thus, heavier growth of plants or algae will have a proportionally higher influence on DO swings each day. Warmer water can hold less oxygen, so summer months tend to see lower DO levels, while colder months will see higher DO levels. Bacterial decay of detritus and other organic matter in sediments consumes oxygen, so DO levels are often lowest at the bottom of the water column, especially when a lake stratifies. The most beneficial method for evaluating DO in lakes is to measure vertical profiles in the water column from surface to bottom, along with water temperature (as an explanatory parameter) to discern the amount of oxygen being consumed towards the bottom of the water column due to degradation of organic matter in sediments. By doing this, we can determine the amount of oxygen available to fish and the fraction of the water column that is habitable by fish during warmer summer months. DO of less than 2 mg/L is considered hypoxic (i.e., low or depleted oxygen), while DO between 3 and 5 mg/L can be stressful for sensitive organisms including larger fish, and DO greater than 5 mg/L is considered generally healthy. DO greater than 8 mg/L is considered optimum for aquatic organism health. However, super-saturated DO levels can be attributed to cyanobacteria blooms and are not necessarily representative of stable conditions.

Results:

At the time of sample and data collection, DO levels ranged from 9.68 mg/L (highest value measured) at the surface to 2.44 mg/L (lowest value measured) at the bottom (Table 3 and Figure 3). These measurements were collected between 9:41 AM at Site 1 through about 11:33 AM at Site 3. The lower DO concentrations at depth demonstrate stratification of the water column which is a normal phenomenon for lakes. Temperature profiles ranged from about 22.8 degrees Celsius at the surface, down to about 20.7 degrees Celsius at the bottom of the water column, again indicating stratification at these open water sites at the time sampled (Table 2 and Figure 2). DO in the groundwater outflow was 3.12 mg/L at 14 degrees Celsius. Thus, the groundwater is more oxygen-depleted when it enters the lake, but it does not appear to be negatively influencing lake-wide DO since it is a relatively small volume being added daily compared to the volume of the lake. Overall, these data indicate that at the time of collection, DO levels were generally good and supportive for aquatic organisms in the upper 80%+ of the water column, with some slight oxygen depletion occurring at the bottom.

3.3 Conductivity

Conductivity is a measure of water's ability to pass an electrical charge. This is directly related to the concentration of dissolved ions in the water (e.g., calcium, magnesium, sodium, potassium, sulfate, and chloride). Conductivity is measured in $\mu\text{S}/\text{cm}$. In freshwater bodies, a conductivity less than 100 $\mu\text{S}/\text{cm}$ is considered low, between 100 and 1,000 $\mu\text{S}/\text{cm}$ is considered normal for freshwaters, and greater than 1,000 $\mu\text{S}/\text{cm}$ may not be tolerable for fish and aquatic invertebrates.

Results:

Conductivity ranged from about 235.2 $\mu\text{S}/\text{cm}$ at the water surface, up to 506.9 $\mu\text{S}/\text{cm}$ at the bottom of the water column (Table 4 and Figure 4). Conductivity in the groundwater outflow was 457.2 $\mu\text{S}/\text{cm}$. All measurements were within normal range for freshwater.

3.4 Phycocyanin

Phycocyanin is a blue-colored pigment produced in cyanobacteria for photosynthesis. This pigment is mostly unique to cyanobacteria, so it can often be used as a screening-level measurement for detection of these organisms in water. While chlorophyll-*a* (pigment discussed below) is a more standardized metric for algal productivity in water, phycocyanin measurements are useful to discern the relative proportion of cyanobacteria within an algal assemblage. These data can be used to prompt further investigation with manual water sampling, algal identification and enumeration, and toxin testing where appropriate.

Results:

Phycocyanin fluorescence values ranged from about 0.27 RFUs at the water surface up to about 12.22 RFUs at the bottom of the water column among the three open water sites (Table 5

and Figure 5). Higher levels in bottom waters at all 3 open water sites suggest algal growth may be occurring near the sediment layer. Phycocyanin was 0.01 RFUs in the groundwater outflow. Overall, all levels were relatively low which would indicate low levels of cyanobacteria in the open water sites at Bass Lake.

3.5 Chlorophyll-*a*

Chlorophyll-*a* is a pigment produced by plants and algae that is used to conduct photosynthesis. Measurements in water provide an estimate of the amount of planktonic (i.e., free-floating) algae and cyanobacteria present and can serve as a metric of productivity in water at that point in time. Chlorophyll-*a* measurements are used as a core metric in determining trophic state of lakes, along with nutrient levels. For example, oligotrophic (low productivity systems) lakes generally have chlorophyll-*a* levels of less than 3 µg/L, while hypereutrophic systems may routinely experience chlorophyll-*a* levels of greater than 40 µg/L. It is important to note that these designations should not be derived from one-time measurements, but rather trends over time. For example, a lake may experience a temporary algal bloom and experience high chlorophyll-*a* levels but does not technically fall under the definition of a hypereutrophic lake. Thus, it is important to use this measurement as a line of evidence in tandem with other water quality parameters.

Results:

At the time of sampling, the YSI ProDSS Sonde was reporting negative values for chlorophyll-*a*. This may have been related to an equipment issue with the optical channel, but we have been investigating and will have the error corrected for the next sampling event. As a result of this issue, all values are reported as zero for the vertical profiles of chlorophyll-*a* in the water column (Table 6). The actual concentration of chlorophyll-*a* (when extracted and analyzed in the lab) ranged from < 8 to 11.8 µg/L (Table 7). These concentrations have declined slightly relative to the previous sampling event in May of 2026.

3.6 Total Phosphorus

Phosphorus is an essential nutrient often correlating with algae growth. Total phosphorus (TP) is the measure of all phosphorus in a sample, reported in µg/L, and includes phosphorus that is readily available for algae and plant growth, as well as phosphorus that has potential to become available. The total phosphorus concentration in an aquatic system is often correlated with the trophic state of water. For example, < 12 µg/L is considered oligotrophic and is often characterized by clear water with little plant or algae growth; 12-24 µg/L is considered mesotrophic and is characterized by moderately clear water with some algae; 25-96 µg/L is considered eutrophic and is characterized by productive waters with algae and plants; > 96 µg/L is considered hypereutrophic (highly productive) and is characterized by frequent algal blooms. Again, these ranges are generalized, and site-specific data should be interpreted in context with other water quality data and lake assessments of productivity. It is helpful to measure

phosphorus in both the surface water and bottom of the water column to discern if there is evidence of phosphorus loading into the water column from sediments. Evidence of this would include significantly higher phosphorus concentrations at the bottom of the water column, especially when a lake is stratified.

Results:

Total phosphorus concentrations ranged from 32 to 40 µg/L in the surface water (Table 7) among Sites 1-3. In the bottom water, concentrations of total phosphorus ranged from < 10 µg/L to 26 µg/L (Table 8). The pump house water had a total phosphorus concentration of 15 µg/L (Table 7). Based on these data, surficial phosphorus levels were in the eutrophic range, and the groundwater is not contributing appreciable phosphorus to the lake. When looking at dissolved oxygen and temperature profiles in addition to bottom water phosphorus levels, these various lines of evidence indicate that while the water column is starting to show declining dissolved oxygen concentrations at the bottom, there is no evidence of significant phosphorus loading from sediments to the water column currently.

3.7 Soluble Reactive Phosphorus

Soluble reactive phosphorus (SRP) is the form of phosphorus in water that is dissolved in water and immediately available for uptake by plants and algae to use for growth. Most of SRP consists of orthophosphate, which is an inorganic form of phosphorus and is the simplest, most biologically available. Since SRP is readily usable, measurement of this parameter can help inform whether there is potential risk for rapid growth of cyanobacteria and algae, as well as reduced water clarity. While there are no universally adopted ranges of SRP that are considered normal/healthy since this form fluctuates often, it is helpful when interpreted in context with total phosphorus concentrations and other water quality parameters. Measurement of SRP is most beneficial in bottom water samples as another line of evidence to discern if lake sediments are a major source of phosphorus release, transformation, and recycling. Surface water dynamics can mask SRP since there is a higher chance for algae in the water to immediately take up the phosphorus, thus this measurement is not as useful in surface water.

Results:

SRP concentrations in bottom waters were all less than the detection limit of 10 µg/L among the three sites sampled (Table 8). This is a good sign that at the time of sampling, there were no appreciable amounts of phosphorus being released from sediments into the bottom water.

3.8 Total Nitrogen

Nitrogen is an essential nutrient that can enhance growth of algae and plants, measured as total nitrogen (TN) in mg/L. Total nitrogen is the combined amount of all forms of inorganic nitrogen (ammonia, nitrite, nitrate) and organic nitrogen (forms found in animal and plant

matter). Nitrogen is usually not a limiting nutrient in aquatic systems but is often compared to total phosphorus levels which can inform whether a lake is more nitrogen-limited or phosphorus-limited. In general, nitrogen levels less than 0.3 mg/L are consistent with nutrient-poor waters, levels between 0.3 and 1 mg/L are considered moderate, levels between 1-2 mg/L are considered a potential eutrophication risk (when interpreted along with phosphorus), and levels greater than 2 mg/L often correlate with highly nutrient enriched systems.

Results:

Total nitrogen concentrations in surface water samples ranged from 1.76 to 2.05 mg/L among the open water sites, while the groundwater had a concentration of 1.12 mg/L (Table 7). When interpreted alongside total phosphorus data, these nitrogen levels are slightly elevated, but common for lakes in the Midwest.

3.9 Total Kjeldahl Nitrogen

Total Kjeldahl Nitrogen (TKN) is a measurement of the sum of organic-N and ammonia-N in water. TKN represents much of the nitrogen tied up in organic matter, which can include nitrogen in algae, plants, and organic debris, as well as in the ammonia form. High TKN levels may provide evidence of runoff containing manure or fertilizers, wastewater influence, decaying organic matter, or dense algae/aquatic vegetation. Organic-N and ammonia-N can both be transformed by bacteria over time into other forms of nitrogen, providing insight into the nitrogen cycle in the lake sampled.

Results:

TKN concentrations ranged from 1.27 to 1.96 mg/L (Table 7) in surface water among the open water sites. The TKN in the groundwater was 1.05 mg/L (Table 7). TKN was the most abundant form of nitrogen in the samples, indicating that there was very little nitrate and nitrite in the water and most of the nitrogen was either in organic form or as ammonia. This means that the nitrogen pool in the lake is currently dominated by algal biomass, dissolved organic nitrogen, particulate organic matter, and reduced nitrogen forms which is common for biologically active lakes.

3.10 Secchi Depth

Secchi depth is a simple measurement for water clarity in a lake. It is determined by using a piece of equipment called a Secchi disc which is a circular disc painted in alternating black and white quadrants, attached to a rope that is lowered down into the water. Secchi depth is defined as the depth of water at which the disc disappears from view as it is lowered. Therefore, a lower Secchi depth indicates highly turbid or algae-dense water, while a higher Secchi depth indicates clearer water. In general, Secchi depths of less than 1 meter (3.3 feet) indicate eutrophic or hypereutrophic conditions, while depths of greater than 6 meters (19.7 feet)

would be indicative of clear water and low productivity systems. However, these ranges vary by region.

Results:

Secchi depth was approximately 2.5-3.3 feet among the three open water sites on the day of sampling (Table 9). Analysts who have routinely been to Bass Lake for other work prior to this visit also noted the water was relatively green compared to other visits to the lake. These Secchi depths measured on June 25, 2026 were slightly greater than those measured in May in the same areas, noting an improvement in water clarity.

3.11 Algal Analysis

The purpose for algal analysis at Bass Lake was to discern the types and relative densities of planktonic cyanobacteria and/or algae. Surficial water samples were collected from five sites within the lake closer to the shoreline. Planktonic algae (especially cyanobacteria) are more likely to be found in shallow, stagnant, and wind-blown shoreline areas in lakes, especially in larger lakes. Therefore, it was important to analyze sites closer to shoreline for algal densities to ensure that if growth was present, it was detected. Sampling only in open water sites could result in false negatives. Samples were analyzed in the laboratory using a light microscope and standardized enumeration methods.

Results:

Several genera of planktonic cyanobacteria (i.e., blue-green algae) were detected in samples collected among the five sampling sites at high densities ranging from 342,880 to 444,840 cells/mL (Table 10). The dominant genus detected at all sites was still *Aphanocapsa* sp., which is a colony-forming type of cyanobacteria, not well established as a cyanotoxin producer. However, overall densities of this genus (and the assemblages as a total) were lower than the prior sampling event. Other genera detected included *Aphanizomenon* sp., *Limnothrix* sp., *Merismopedia* sp., and *Microcystis* sp. These four genera of cyanobacteria are potential toxin producers.

While toxin testing was not performed on these samples, our laboratory can offer that testing if Bass Lake is interested in future sampling events. We recommend keeping dogs out of water that appears dense green/blue-green along lake shorelines. Similarly, if these signs of dense cyanobacteria are present, we also recommend that people not swim or otherwise come in contact with the water. For more information on harmful algal bloom guidance in Indiana, please refer to IDEM's webpage: <https://www.in.gov/idem/algae/indiana-reservoir-and-lake-sampling-update/>.

Since algal assemblages were dominated by cyanobacteria, this would indicate potentially higher levels of phosphorus around the lake shoreline. Higher phosphorus concentrations around shoreline areas can result from runoff of nutrients due to rain events

(especially in springtime), potential groundwater runoff from leaky septic tanks, or from the release of phosphorus that can occur following invasive vegetation treatments as the plant material gets degraded by bacteria.

Table 1. pH measurements (reported in standard units) collected with depth on June 25, 2026. Pumphouse outflow water was collected in a bucket for measurement with sonde.

Depth (m)	Site 1	Site 2	Site 3	Pumphouse
0	8.67	8.68	8.80	7.40
1	8.65	8.72	8.80	
2	8.65	8.71	8.81	
3	8.63	8.68	8.76	
4	8.41	8.24	8.73	
5	8.31	8.12	8.48	
6	6.35	8.06	6.41	
7	-	8.01	-	

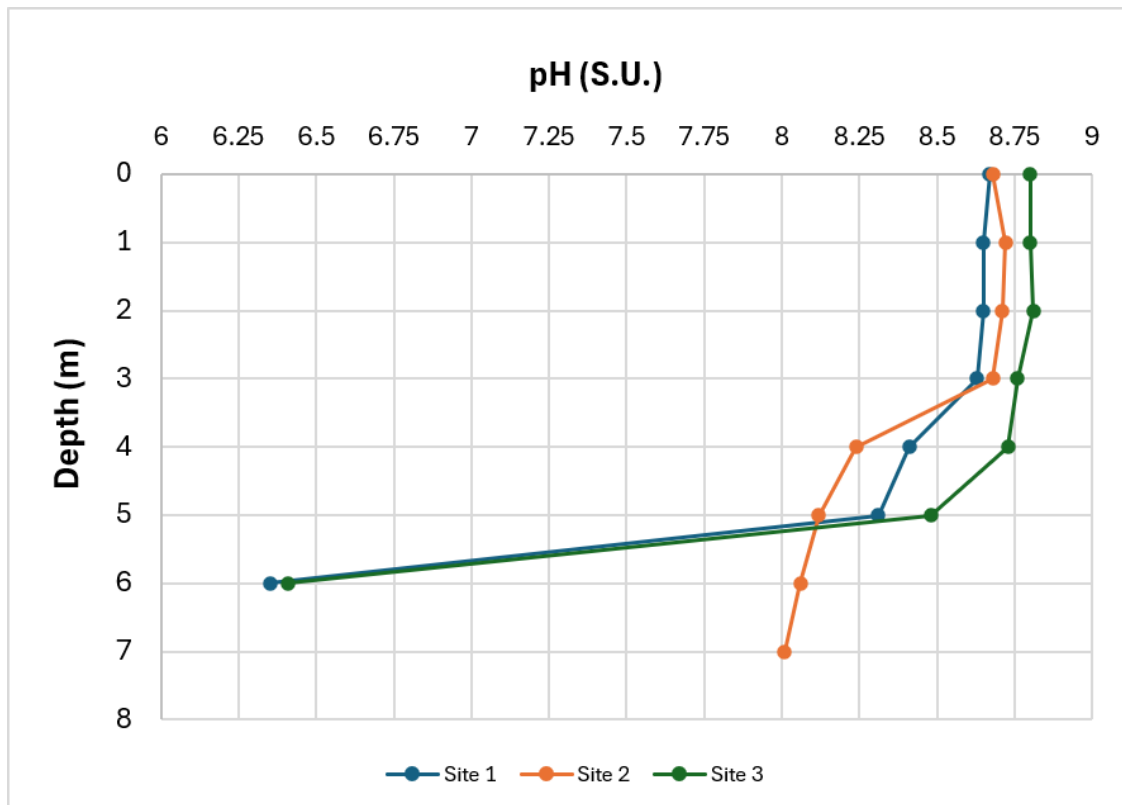


Figure 1. Graph of pH measurements (reported in standard units) collected with depth on June 25, 2026. Pumphouse measurement only reported in Table 1.

Table 2. Temperature measurements (reported in degrees Celsius) collected with depth on June 25, 2026. Pumphouse outflow water was collected in a bucket for measurement with sonde.

Depth (m)	Site 1	Site 2	Site 3	Pumphouse
0	22.0	22.8	22.4	13.9
1	21.8	22.1	22.2	
2	21.6	21.9	21.9	
3	21.5	21.6	21.7	
4	21.3	21.4	21.5	
5	21.1	21.3	21.3	
6	20.7	21.2	20.8	
7	-	21.2	-	

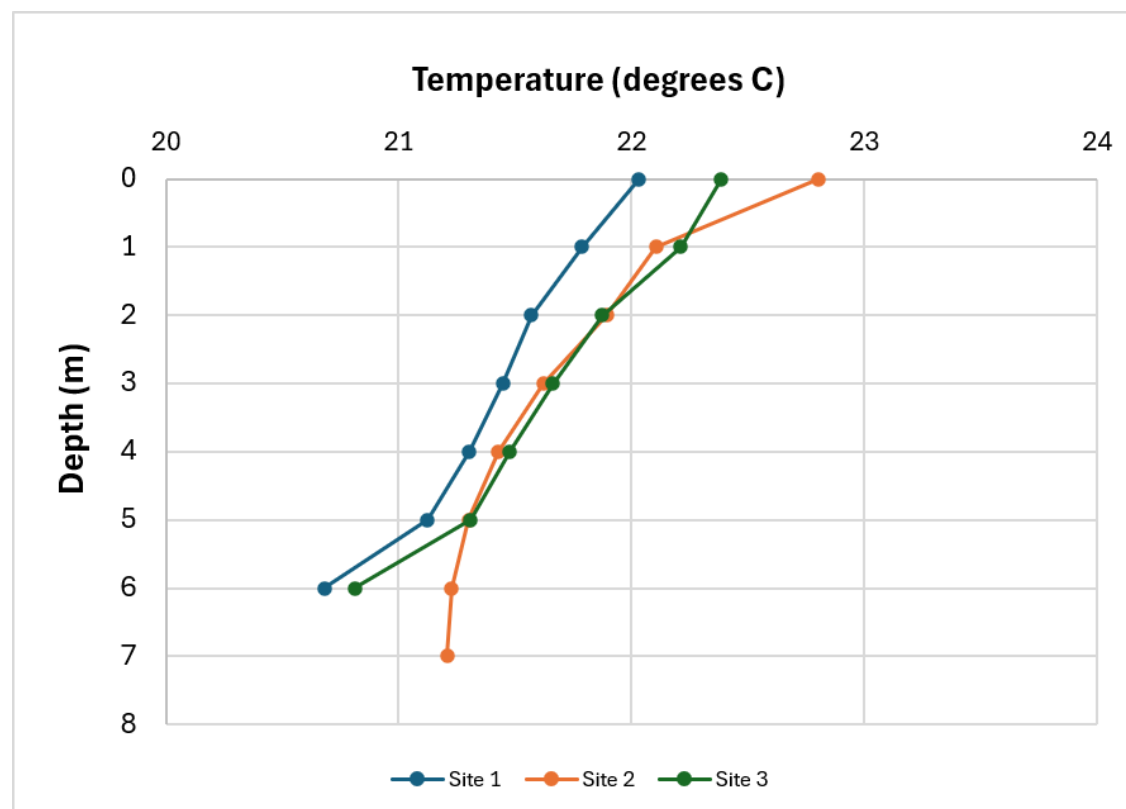


Figure 2. Graph of temperature measurements (reported in degrees Celsius) collected with depth on June 25, 2026. Pumphouse measurement reported in Table 2.

Table 3. Dissolved oxygen measurements (reported in mg/L) collected with depth on June 25, 2026. Pumphouse outflow water was collected in a bucket for measurement with sonde.

Depth (m)	Site 1	Site 2	Site 3	Pumphouse
0	9.02	9.15	9.68	3.12
1	8.98	9.42	9.69	
2	8.81	9.32	9.76	
3	8.80	9.15	9.45	
4	8.00	7.84	9.23	
5	7.52	7.15	8.04	
6	2.83	6.39	2.44	
7	-	6.08	-	

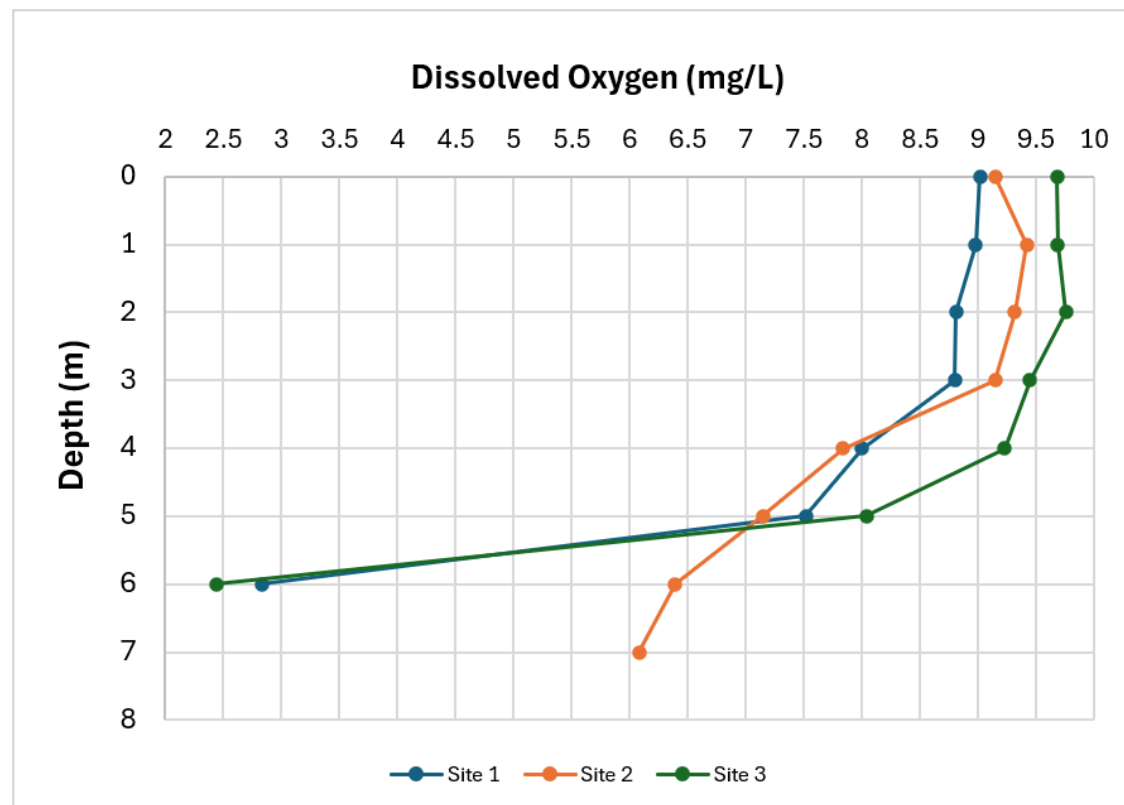


Figure 3. Graph of dissolved oxygen measurements (reported in mg/L) collected with depth on June 25, 2026. Pumphouse measurement reported in Table 3.

Table 4. Conductivity measurements (reported in $\mu\text{S}/\text{cm}$) collected with depth on June 25, 2026. Pumphouse outflow water was collected in a bucket for measurement with sonde.

Depth (m)	Site 1	Site 2	Site 3	Pumphouse
0	231.0	235.2	233.1	457.2
1	229.9	231.6	231.8	
2	228.2	230.8	229.8	
3	228.0	229.9	229.0	
4	228.3	232.0	227.7	
5	228.7	233.3	230.4	
6	388.3	234.1	506.9	
7	-	234.3	-	

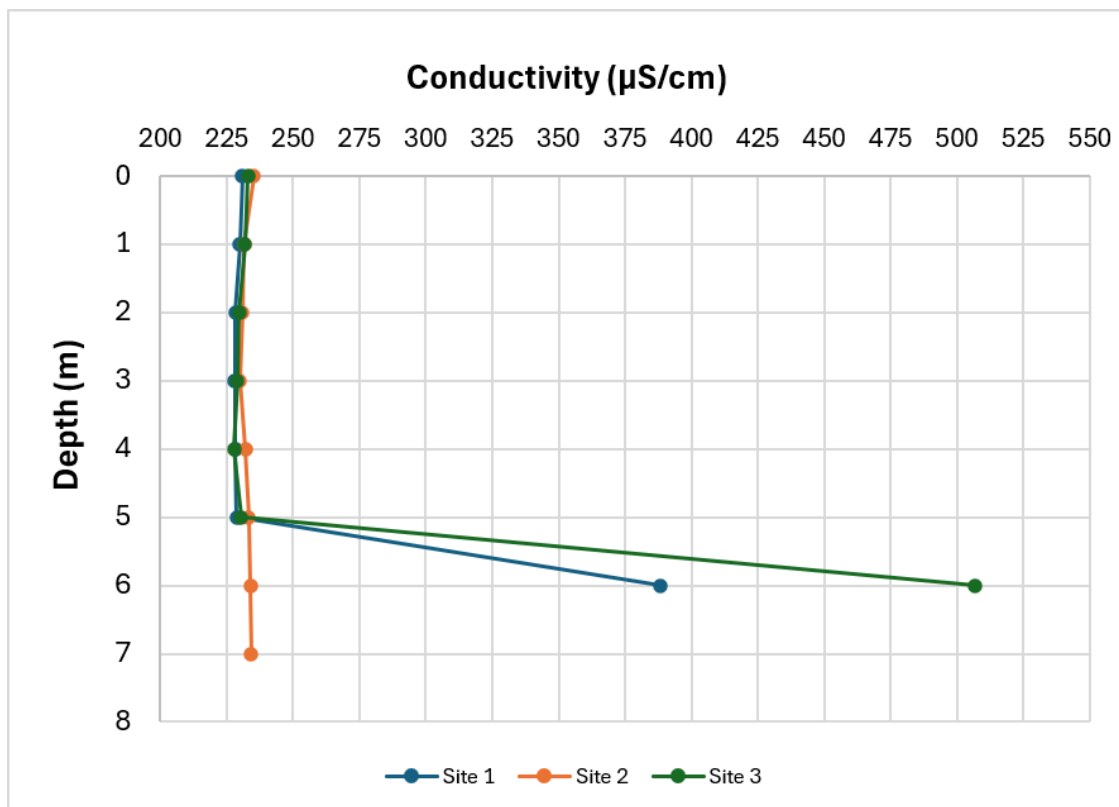


Figure 4. Graph of conductivity measurements (reported in $\mu\text{S}/\text{cm}$) collected with depth on June 25, 2026. Pumphouse measurement reported in Table 4.

Table 5. Phycocyanin measurements (reported in relative fluorescence units; RFUs) collected with depth on June 25, 2026. Pumphouse outflow water was collected in a bucket for measurement with sonde.

Depth (m)	Site 1	Site 2	Site 3	Pumphouse
0	0.35	0.32	0.27	0.01
1	0.54	0.42	0.32	
2	0.56	0.48	0.41	
3	0.51	0.64	0.54	
4	0.95	0.96	0.59	
5	1.35	1.33	1.01	
6	12.22	1.84	7.31	
7		2.21		

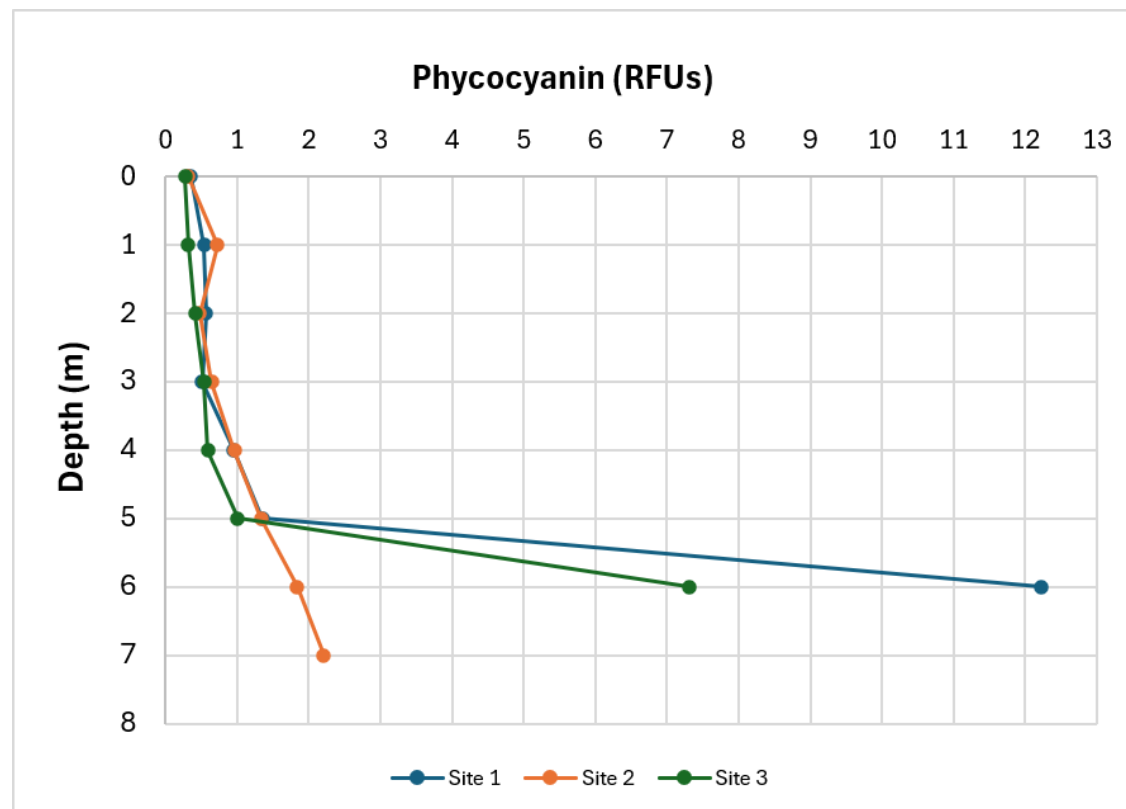


Figure 5. Graph of phycocyanin measurements (reported in relative fluorescence units; RFUs) collected with depth on June 25, 2026. Pumphouse measurement reported in Table 5.

Table 6. Chlorophyll-*a* measurements (reported in relative fluorescence units; RFUs) collected with depth on June 25, 2026. Pumphouse outflow water was collected in a bucket for measurement with sonde.

Note: YSI ProDSS sonde reported negative values for chlorophyll-*a*, so no graph was created for these data, and all values are reported as zero here. We are investigating a potential equipment issue related to the optical channel and will ensure all necessary repairs are made before the next sampling visit.

Depth (m)	Site 1	Site 2	Site 3	Pumphouse
0	0	0	0	0
1	0	0	0	
2	0	0	0	
3	0	0	0	
4	0	0	0	
5	0	0	0	
6	0	0	0	
7	0	0	0	

Table 7. Nutrient and pigment data from surface samples collected at Bass Lake on June 25, 2026

Sample ID	Total Phosphorus (µg/L)	Total Nitrogen (mg/L)	Total Kjeldahl Nitrogen (mg/L)	Chlorophyll-a (µg/L)
Site 1	32	2.05	1.96	8.7
Site 2	34	1.84	1.77	< 8
Site 3	40	1.76	1.27	11.8
Pumphouse	15	1.12	1.05	< 8

Table 8. Water quality data from bottom samples and pumphouse sample collected at Bass Lake on June 25, 2026

Sample ID	Total Phosphorus (µg/L)	Soluble Reactive Phosphorus (µg/L)
Site 1	21	< 10
Site 2	< 10 (6)*	< 10
Site 3	26	< 10
Pumphouse	15	< 10

* Result was below reporting limit. Estimated concentration in parentheses.

Table 9. Secchi depth measurements collected at Bass Lake on June 25, 2026

Sample Site	Secchi Depth (feet)
Site 1	2.5
Site 2	3.3
Site 3	3.3

Table 10. Algal identification and enumeration results

Site ID	Genus (Taxonomic Class)	Cell Density by Genus (cells/mL)	Total Cell Density (cells/mL)
Site 1	<i>Aphanizomenon sp.</i> (Cyanophyceae)	480	342,880
	<i>Aphanocapsa sp.</i> (Cyanophyceae)	263,710	
	<i>Limnothrix sp.</i> (Cyanophyceae)	76,230	
	<i>Microcystis sp.</i> (Cyanophyceae)	2,460	
Site 2	<i>Aphanocapsa sp.</i> (Cyanophyceae)	318,770	378,650
	<i>Limnothrix sp.</i> (Cyanophyceae)	53,520	
	<i>Merismopedia sp.</i> (Cyanophyceae)	3,150	
	<i>Microcystis sp.</i> (Cyanophyceae)	3,210	
Site 3	<i>Aphanocapsa sp.</i> (Cyanophyceae)	238,700	394,710
	<i>Limnothrix sp.</i> (Cyanophyceae)	153,740	
	<i>Merismopedia sp.</i> (Cyanophyceae)	1,620	
	<i>Microcystis sp.</i> (Cyanophyceae)	650	
Site 4	<i>Aphanocapsa sp.</i> (Cyanophyceae)	360,080	441,040
	<i>Limnothrix sp.</i> (Cyanophyceae)	71,240	
	<i>Merismopedia sp.</i> (Cyanophyceae)	9,170	
	<i>Microcystis sp.</i> (Cyanophyceae)	550	
Site 5	<i>Aphanizomenon sp.</i> (Cyanophyceae)	1,060	444,840
	<i>Aphanocapsa sp.</i> (Cyanophyceae)	387,530	
	<i>Limnothrix sp.</i> (Cyanophyceae)	56,250	

Note: Cyanophyceae is the taxonomic class that includes cyanobacteria (i.e., “blue-green algae”)

All data presented in this report met QA/QC criteria unless otherwise specified here.

Field Notes and Data Sheet
Bass Lake; Knox IN

Sample Date: 6/25/26

Analyst: Malena Ryan

Notes on weather: Sunny and blue skies, minimal wind compared to last sampling; around 75 degrees air temp.

Approximate time of sample/data collection:

Open Water Site 1: 9:15AM CST

Open Water Site 2: 10:40AM CST

Open Water Site 3: 11:28AM CST

Pumphouse: 11:20AM CST

Algae Site 1: 9:10AM CST

Algae Site 2: 10:15AM CST

Algae Site 3: 10:30AM CST

Algae Site 4: 11:40AM CST

Algae Site 5: 11:55AM CST

Secchi Depth Measurements:

Site ID	Secchi Depth (ft)
Open Water Site 1	2.5
Open Water Site 2	3.3 ft
Open Water Site 3	3.3 ft

Observations: Water is still very green; very little plant growth